Cancer Prevention With Dehydroepiandrosterone and Non-Androgenic Structural Analogs

Arthur G. Schwartz, PhD and Laura L. Pashko, PhD

Fels Institute for Cancer Research and Molecular Biology and Department of Microbiology and Immunology, Temple University School of Medicine, Philadelphia, PA 19140

Abstract There is increasing evidence that the adrenocortical steroid, dehydroepiandrosterone (DHEA), is an important mammalian hormone. Administration of DHEA to laboratory mice and rats inhibits development of experimental tumors of the breast, lung, colon, liver, skin and lymphatic tissue. In the two-stage skin tumorigenesis model in mice, DHEA treatment inhibits tumor initiation, as well as tumor promoter-induced epidermal hyperplasia and promotion of papillomas. There is much evidence that DHEA produces its antiproliferative and tumor preventive effects by inhibiting glucose-6-phosphate dehydrogenase and the pentose phosphate pathway. This pathway is an important source of NADPH, a critical reductant for many biochemical reactions that generate oxygen free radicals, which may act as second messengers in stimulating hyperplasia. The therapeutic use of DHEA in humans may be limited by its sex hormonal side effects. DHEA is metabolized *in vivo* to both testosterone and estrone, producing both androgenic and estrogenic effects in laboratory animals. We have developed a synthetic steroid, 16 α -fluoro-5-androsten-17-one, which does not demonstrate the androgenic or estrogenic activity of DHEA, yet retains the antiproliferative and cancer preventive activity of the native steroid. **© 1995 Wiley-Liss, Inc.**

Key words: Adrenal gland, dehydroepiandrosterone, glucose-6-phosphate dehydrogenase

Reducing the food intake of laboratory mice and rats produces the most marked cancer preventive effect of any known regimen—it inhibits the development of spontaneous [1,2], chemically induced [1,3], and radiation-induced [4,5] tumors. Not only does underfeeding inhibit tumorigenesis, it also retards the rate of development of numerous age-related physiologic and pathologic changes and apparently retards the rate of aging [6]. Numerous theories have been proposed to account for the mechanism by which food restriction exerts its remarkable beneficial effects, but no satisfactory explanation has emerged [6].

© 1995 Wiley-Liss, Inc.

DEHYDROEPIANDROSTERONE

Over the past several years, this laboratory and others have demonstrated that administration of the adrenocortical steroid, dehydroepiandrosterone (DHEA), to laboratory mice and rats inhibits tumor development in the breast [7], lung [8], skin [9], liver [10], colon [11] and lymphatic tissue [12]. DHEA treatment of mice and rats also reduces the rate of weight gain [13,14]. This reduction of weight gain is not simply a result of impaired food intake but an apparent consequence of the stimulation of thermogenesis with a concomitant reduction in metabolic efficiency [15]. In addition to protecting animals against cancer, DHEA treatment produces many of the beneficial effects of food restriction, including an inhibition of experimentally induced atherosclerosis [16,17], suppression of age-related

Address correspondence to Arthur G. Schwartz, PhD, Temple University School of Medicine, Fels Institute for Cancer Research, 3420 North Broad Street, Philadelphia, PA 19140.

proteinuria development [18], and inhibition of autoimmune disease development [19]. The antiweight effect of DHEA, as well as its ability to produce many of the beneficial effects of food restriction, led us to hypothesize that elevated levels of DHEA in food-restricted rodents might partly mediate the tumor-suppressing and ageretarding effects of underfeeding [20].

We found previously that 12-O-tetradecanoylphorbol-13-acetate (TPA) stimulation of mouse epidermal hyperplasia (as measured by an increase in [³H]thymidine incorporation or an increase in epidermal DNA content of a 2x2 cm² area of skin) is blocked by one to two weeks of food restriction prior to TPA application [21]. This relatively rapid response of the hyperplastic effect of the tumor promoter to underfeeding enables testing of experimental manipulations that might alter the responsiveness of skin tumor promotion to underfeeding.

ROLE OF ADRENOCORTICAL STEROIDS IN MEDIATING TUMOR-SUPPRESSING EFFECT OF UNDERFEEDING

As long ago as 1948, Boutwell *et al.* [22] reported that food restriction of mice appeared to activate adrenocortical activity, as evidenced by thymic involution, a decrease in blood lymphocyte counts, an enhanced gluconeogenesis rate, and an increase in the ratio of adrenal gland weight to body weight. We have found about a two-fold increase in plasma corticosterone levels in food-restricted CD-1 mice (28% less than *ad libitum* fed) after 11 weeks [23].

Our studies with CD-1 mice found that food restriction abolished TPA stimulation of epidermal [³H]thymidine incorporation and markedly suppressed TPA promotion of papillomas. Adrenalectomy of mice prior to initiating food restriction completely reversed the inhibitory effect of underfeeding on TPA stimulated [³H]thymidine incorporation and promotion of tumors, indicating clearly that the adrenal gland plays a critical role in inhibiting tumor promoter-induced hyperplasia and tumor promotion in food-restricted mice [23].

We also found that adrenalectomy completely reverses the tumor-inhibitory effect of food restriction in a lung tumor model. In this study, lung adenomas were induced in male A/J mice by a single oral dose of 0.5 mg of 7,12-dimethylbenz(*a*)anthracene. One week later the mice were either adrenalectomized or sham operated and thereafter fed *ad libitum* or given 27% less food than the *ad libitum*-fed group. Fourteen weeks later the mice were sacrificed and the number of alveolar adenomas counted. Food restriction reduced the number of adenomas four-fold in the sham-operated mice, whereas adrenalectomy completely abolished the effect of food restriction. Adrenalectomy also enhanced the number of alveolar adenomas; both the *ad libitum* and food-restricted adrenalectomized mice had twice the number of adenomas as the *ad libitum* shamoperated mice [unpublished observation].

Two adrenocortical steroids, corticosterone and DHEA, inhibit TPA-stimulated epidermal hyperplasia and TPA promotion of skin tumors in mice [9,24]. We hypothesize that overproduction of these steroids in response to food restriction accounts for the tumor inhibitory effect of underfeeding [25].

EFFECT OF GLUCOCORTICOID RECEPTOR ANTAGONIST

The synthetic steroid mifepristone, RU-486, is a potent progestin and glucocorticoid antagonist [26]. RU-486 has a very high binding affinity for the glucocorticoid receptor (about 31 times that of corticosterone and at least as high as potent agonists such as dexamethasone [27]). When administered *in vivo* to rats (at 10–25 mg/kg, po), RU-486 completely prevents the thymolytic activity of corticosterone and dexamethasone [28].

The availability of RU-486 enables testing of the hypothesis that elevated levels of glucocorticoid steroids contribute to the inhibition by food restriction of TPA-promoted skin tumors. We first determined that orally administered RU-486 reversed the suppression produced by corticosterone treatment in TPA-stimulated epidermal hyperplasia. Hyperplasia was measured by guantitating the epidermal DNA content of a 2x2 cm² section of mouse skin 48 hours after TPA application. RU-486 was administered po as a solution in sesame oil or given in the diet to yield a dose of approximately 25 mg/kg. As shown in Table I, 200 µg of topically applied corticosterone abolished the TPA stimulation in epidermal DNA content, and RU-486 completely reversed the inhibitory effect of corticosterone. This dose of RU-486 also had no effect on inhibition produced by

Treatment Group	DNA Content of 2x2 cm ² of Mouse Epidermis (µg)
Control (no TPA)	73.9 ± 9.1
TPA	132 ± 7.4
TPA + RU-486	138 ± 20.2
TPA + Corticosterone	59.8 ± 8.6
TPA + Corticosterone + RU-486	$129 \pm 12.7^{*}$

TABLE I. Effect of RU-486 Administration on
Corticosterone-induced Suppression in
TPA-Stimulated Epidermal DNA Content

The backs of 6–7 week old CD-1 female mice were shaved, and only those mice showing no hair growth were used. Two days prior to TPA treatment, mice were given Purina 5015 chow or chow containing RU-486 (at a level yielding a dose of 23–24 mg/kg body weight). Two days later, mice were treated topically with 200 µg corticosterone in 0.2 ml acetone, or with acetone vehicle, one hour before TPA treatment, (2 µg in 0.2 ml of acetone). Mice received a second treatment of 200 µg corticosterone or acetone vehicle 24 hours later, and 24 hours after this, the mice were sacrificed. The epidermis was isolated from a 2x2 cm² piece of skin, and the DNA content was determined [38]. Each value is the mean \pm S.D. for five separately treated mice.

* > TPA + Corticosterone group, p < 0.001.

DHEA treatment in TPA-induced epidermal hyperplasia (data not shown).

As mentioned previously, food restriction of CD-1 mice blocks TPA-stimulated epidermal hyperplasia and inhibits TPA promotion of papillomas. We found that RU-486 administration, at doses that completely reversed corticosteroneinduced suppression of TPA-stimulated epidermal hyperplasia, significantly reversed the inhibition produced by eight days of food restriction in TPA-stimulated hyperplasia. We have consistently observed this effect of RU-486, whether the drug is administered po in sesame oil or in the diet, and these data strongly suggest that elevated corticosterone levels in food-restricted mice significantly inhibit TPA-induced epidermal hyperplasia (Table II).

We then determined if RU-486 administration in the diet significantly reversed the suppression produced by food restriction in papilloma development, and much to our surprise, we found no

Table II.	Effect of RU-486 Administration	
on Food	Restriction-induced Inhibition in	
TPA-Stin	nulated Epidermal DNA Content	

Treatment Group	DNA Content of 2x2 cm ² of Mouse Epidermis (µg)
Control	4 9.7 ± 12.3
TPA	138 ± 20.1
TPA + RU-486	151 ± 37.2
Food Restricted	49.6 ± 7.2
Food Restricted + TPA	39.7 ± 9.5
Food Restricted + TPA + RU-486	$121 \pm 23.5^{*}$

Six to seven week old female CD-1 mice were singly housed and given *ad libitum* access to Purina 5015 chow or food-restricted with daily allotments of 62% of the food consumed by the *ad libitum* fed animals. RU-486-treated mice received RU-486 daily in the chow to give a dose of 25 mg/kg body weight. Eight days after initiating food restriction and RU-486 treatment, mice were treated on their shaved backs with 2 µg TPA in 0.2 ml acetone or with acetone vehicle. Forty-eight hours later, the mice were sacrificed and epidermal DNA content of a 2x2 cm² section of mouse skin was determined [38]. Each value is the mean \pm S.D. of five separately treated mice.

* > Food Restricted + TPA group, p < 0.001.

effect of RU-486 treatment on food restrictioninduced suppression in skin tumor formation (Fig. 1). Since it seemed very likely to us that any agent which reversed epidermal hyperplasia suppression induced by food restriction would also reverse tumorigenesis inhibition, we determined the effect of RU-486 administration on TPA-induced hyperplasia following four weeks of food restriction. We found that RU-486 treatment during a four-week period of food restriction had no effect on suppressing TPA-induced hyperplasia produced by underfeeding (Table III).

In humans, RU-486 blocks the inhibitory feedback effect of cortisol on corticotrophin secretion, leading to an increase in plasma cortisol and corticotrophin levels [29]. Thus, it is possible that plasma corticosterone levels were sufficiently elevated in long-term RU-486-treated animals to overcome the RU-486 blockade of glucocorticoid receptors. Alternatively, elevated levels of DHEA

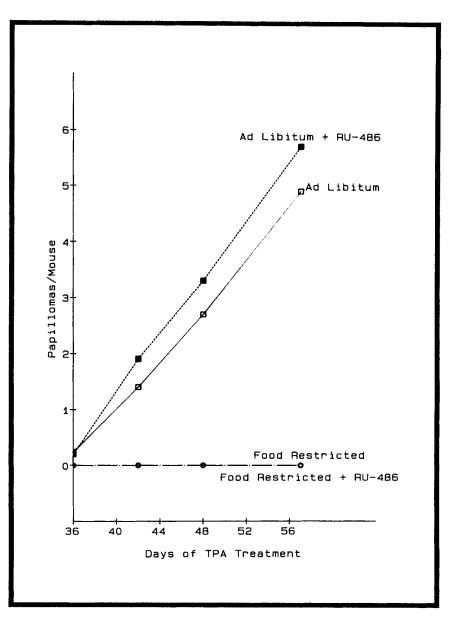


Fig. 1. Effect of RU-486 administration on food restriction-induced suppression in TPA promotion of mouse skin papillomas. Female CD-1 mice were initiated on the shaved backs with 200 nmol 7,12-dimethylbenz(*a*)anthracene in 0.2 ml acetone. Seven days later, the mice were given *ad libitum* access to chow or restricted with daily allotments of 56% of the food consumed by the *ad libitum*-fed mice. One group of *ad libitum*-fed mice and one group of food-restricted mice also received RU-486 in the diet to yield a daily dose of drug of 24 ± 1.5 mg/kg body weight. Eight days after carcinogen initiation, all mice were treated with twice weekly applications of 2 µg TPA in 0.1 ml acetone. Mice were palpated weekly for papillomas, and the mean number of tumors per group was determined. There were 23 mice in each of the RU-486-treated groups, and 25 mice in each of the non-RU-486-treated groups.

Table III. Effect of RU-486 Administration on Food Restriction-induced Inhibition in TPA-Stimulated Epidermal DNA Content (Four Week Food Restriction)

Treatment Group	DNA Content of 2x2 cm ² of Mouse Epidermis (µg)
Control	53.3 ± 12.1
TPA	120 ± 20.4
Food Restricted	42.7 ± 5.6
Food Restricted + TPA	43.5 ± 8.5
Food Restricted + TPA + RU-486	39.0 ± 13.8

The experimental conditions were the same as in Table II except that mice were food restricted and treated with RU-486 for four weeks instead of eight days.

or a DHEA-like steroid in the RU-486 foodrestricted mice may contribute to the inhibition of TPA-induced epidermal hyperplasia and TPA promotion of tumors. The inhibition in TPAinduced hyperplasia and tumor promotion produced by DHEA administration, unlike the inhibition produced by corticosterone, is not reversed by RU-486 treatment (unpublished results).

BIOLOGICAL SIGNIFICANCE OF DEHYDROEPIANDROSTERONE

DHEA, along with its sulfate ester, is secreted by the human adrenal cortex in quantities at least as great as all the other adrenocortical steroids combined [30]. There is increasing evidence that this substance may be an important adrenocortical hormone [31]. The plasma level of DHEA sulfate is highest in the second decade, then declines markedly [32]. Epidemiological studies suggest that low plasma levels of this steroid may predispose individuals to develop certain cancers [33,34] and die from cardiovascular disease [35].

DHEA is a potent uncompetitive inhibitor of mammalian glucose-6-phosphate dehydrogenase [36], the rate-controlling enzyme in the pentose phosphate pathway. This pathway is an important source of NADPH as well as ribose 5-phosphate. Inhibition of this pathway by DHEA apparently accounts for the antiproliferative and antitumor promoting actions of the steroid [37,38]. Probably as a result of a reduction in NADPH levels, DHEA, also inhibits mixed-function oxidase activation of chemical carcinogens [39], the NADPH-dependent production of oxygen free radicals from paraquat [40], and the oxidative burst generation of O_2^- [41].

There is increasing evidence that oxygen free radicals contribute to the development of many age-related diseases, including cancer [42], atherosclerosis [43], and the familial autosomal form of amyotrophic lateral sclerosis, as well as other neurodegenerative diseases [44]. Oxygen free radicals, aside from directly damaging DNA and producing mutations [45], may also serve as second messengers stimulating hyperplasia [46]. The importance of oxygen free radicals in tumor promotion is underscored by the marked suppression of TPA promotion of skin tumor formation by treatment with a low molecular weight copper chelate with superoxide dismutase activity [47].

We have postulated that the glucocorticoid steroids suppress inflammation, an important source of oxygen free radicals; DHEA inhibits the pentose phosphate pathway and the generation of NADPH, a critical reductant for reactions generating oxygen free radicals [45,48–50]. Together they may mediate the tumor suppressing and possibly the age-retarding effects of underfeeding [25].

DEHYDROEPIANDROSTERONE ANALOGS

The striking biological effects of DHEA in laboratory animals suggest that this steroid, or specific structural analogs, could become an important class of drug in humans. DHEA was efficacious in treating systemic lupus erythematosus and type II diabetes in early clinical trials [51-53]. However, DHEA has side effects that may limit its use in humans. It is metabolized into both testosterone and estrone and produces estrogenic and androgenic effects in laboratory rats [54]. Pharmacological doses of DHEA given to six postmenopausal women for 28 days produced a marked elevation of plasma testosterone (9-fold) and dihydrotestosterone (20-fold) [55]. DHEA treatment induced insulin resistance in these women and significantly lowered plasma HDL levels, very likely as a result of the androgenic state induced. DHEA is also a peroxisome proliferator; its long-term administration to Fischer rats induces a high incidence of hepatocellular carcinoma [56].

We have developed the synthetic steroid, 16α -fluoro-5-androsten-17-one, which does not demonstrate the androgenic, estrogenic, or peroxisome-proliferating properties of DHEA, yet has retained the antiproliferative, cancer preventive, anti-obesity and anti-diabetic actions of the native steroid [54,57,58]. Compounds such as 16α -fluoro-5-androsten-17-one may become important therapeutic agents in humans.

ACKNOWLEDGMENTS

This work was supported by NIH grants CA 52500 and CA 12227. All animal care in these experiments was in accordance with the guidelines of the Institutional Animal Care and Use Committee of Temple University.

The authors thank Dr. Etienne-Emile Baulieu and Roussel Uclaf for kindly providing us with RU-486 for these experiments.

REFERENCES

- 1. Tannenbaum A, Silverstone H: Nutrition in relation to cancer. Adv Cancer Res 1:451–501, 1953.
- Shimokawa I, Yu BP, Masoro EJ: Influence of diet on fatal neoplastic disease in male Fischer 344 rats. J Gerontol 46:228–232, 1991.
- Klurfeld DM, Weber MM, Kritchevsky D: Inhibition of chemically induced mammary and colon tumor promotion by caloric restriction in rats fed increased dietary fat. Cancer Res 47:2759–2762, 1987.
- Gross L, Dreyfuss Y: Inhibition of the development of radiation-induced leukemia in mice by reduction of food intake. Proc Natl Acad Sci USA 83:7928–7931, 1986.
- Gross L, Dreyfuss Y: Reduction in the incidence of radiation-induced tumors in rats after restriction of food intake. Proc Natl Acad Sci USA 81:7596–7598, 1984.
- Masoro EJ: Nutrition as a modulator of the aging process. Physiologist 27:98–101, 1984.
- Schwartz AG: Inhibition of spontaneous breast cancer formation in C3H-A^{vy}/a mice by long-term treatment with dehydroepiandrosterone. Cancer Res 39: 1129–1132, 1979.
- Schwartz AG, Tannen RH: Inhibition of 7,12-dimethyl(*a*)anthracene- and urethan-induced lung tumor formation in A/J mice by long-term treatment with dehydroepiandrosterone. Carcinogenesis 2:1335–1338, 1981.

- Pashko LL, Rovito RJ, Williams JR, Sobel EL, Schwartz AG: Dehydroepiandrosterone (DHEA) and 3β-methylandrost-5-en-17-one: Inhibitors of 7,12-dimethylbenz(a)anthracene (DMBA)-initiated and 12-Otetradecanoylphorbol-13-acetate (TPA)-promoted skin papilloma formation in mice. Carcinogenesis 5:463– 466, 1984.
- 10. Weber E, Moore MA, Bannasch P: Phenotypic modulation of hepatocarcinogenesis and reduction in *N*nitroso-morpholine-induced hemangiosarcoma and adrenal lesion development in Sprague-Dawley rats by dehydroepiandrosterone. Carcinogenesis 9:1191– 1195,1988.
- 11. Nyce JW, Magee PN, Hard GC, Schwartz AG: Inhibition of 1,2-dimethylhydrazine-induced colon tumorigenesis in Balb/c mice by dehydroepiandrosterone. Carcinogenesis 5:57–62, 1984.
- Mei JM, Hursting SD, Phang JM: Chemoprevention of spontaneous tumorigenesis in p53 knocknout mice. Proc Am Assoc Cancer Res, 1995 (in press).
- Yen TT, Allan JV, Pearson DV, Acton JM, Greenberg M: Prevention of obesity in A^{vy}/a mice by dehydroepiandrosterone. Lipids 12:401–413, 1977.
- Cleary MP, Fox N, Lazin B, Billheimer JT: A comparison of the effects of dehydroepiandrosterone treatment to *ad libitum* and pair feeding in the obese Zucker rat. Nutr Res 5:1247–1257, 1985.
- Lardy H, Su C-Y, Kneer N, Wielgus S: Dehydroepiandrosterone induces enzymes that permit thermogenesis and decrease metabolic efficiency. In Lardy H, Stratman F (eds): "Hormones, Thermogenesis, and Obesity." New York: Elsevier, 1988, pp 415–426.
- Gordon GB, Bush DE, Weisman HF: Reduction of atherosclerosis by administration of dehydroepiandrosterone. J Clin Invest 82:712–720, 1988.
- Eich DM, Nestler JE, Johnson DE, Dworkin GH, Ko D, Wechsler AS, Hess ML: Inhibition of accelerated atherosclerosis with dehydroepiandrosterone in the heterotropic rabbit model of cardiac transplantation. Circulation 87:261–269, 1993.
- Pashko LL, Fairman DK, Schwartz AG: Inhibition of proteinuria development in aging Sprague-Dawley rats and C57BL/6 mice by long-term treatment with dehydroepiandrosterone. J Gerontol 41:433–438, 1986.
- Lucas JA, Ahmed SA, Casey JL, MacDonald PC: Prevention of autoantibody formation and prolonged survival in New Zealand Black/New Zealand White female mice fed dehydroepiandrosterone. J Clin Invest 75:2091–2093, 1985.
- Schwartz AG, Fairman DK, Pashko LL: The biological significance of dehydroepiandrosterone. In Kalimi M, Regelson W (eds): "The Biologic Role of Dehydroepiandrosterone." West Berlin: Walter de Gruyter Press, 1990, pp 7–12.
- Schwartz AG, Pashko LL: Food restriction inhibits [³H]7,12-dimethylbenz[*a*]anthracene binding to mouse skin DNA and tetradecanoylphorbol-13-acetate stimulation of epidermal [³H]thymidine incorporation. Anticancer Res 6:1279–1282, 1986.
- 22. Boutwell RK, Brush MK, Rusch HP: Some physiolo-

gical effects associated with chronic caloric restriction. Am J Physiol 154:517-524, 1948.

- Pashko LL, Schwartz AG: Reversal of food restriction-induced inhibition of mouse skin tumor promotion by adrenalectomy. Carcinogenesis 13:1925–1928, 1992.
- Schwarz JA, Viaje A, Slaga TJ, Yuspa SH, Hennings H, Lichti U: Fluocinolone acetonide: A potent inhibitor of mouse skin tumor promotion and epidermal DNA synthesis. Chem Biol Interact 17:331–347, 1977.
- Schwartz AG, Pashko LL: Role of adrenocortical steroids in mediating cancer-preventive and ageretarding effects of food restriction in laboratory rodents. J Gerontol 49:B37–B41, 1994.
- Spitz IM, Bardin CW: Mifepristone (RU-486)—a modulator of progestin and glucocorticoid action. N Engl J Med 329:404–412, 1993.
- Moguilewsky M, Philibert D: Biochemical profile of RU-486. In Segal SJ, Baulieu EE (eds): "The Antiprogestin Steroid RU-486 and Human Fertility Control." New York: Plenum Press, 1985, pp 87–97.
- Philibert D, Moguilewsky M, Mary I, Lecaque D, Tournemine C, Seechi J, Deraedt R: Pharmacological profile of RU-486 in animals. In Segal SJ, Baulieu EE (eds): "The Antiprogestin Steroid RU-486 and Human Fertility Control." New York: Plenum Press, 1985, pp 49–68.
- Gaillard RC, Riondel A, Muller AF, Herrmann W, Baulieu EE: RU-486: A steroid with antiglucocorticoid activity that only disinhibits the human pituitary-adrenal system at a specific time of day. Proc Natl Acad Sci USA 81:3879–3882, 1984.
- Gregerman RI, Bierman EL: Aging and hormones. In Williams RH (ed): "Textbook of Endocrinology." Philadelphia: W.B. Saunders, 1981, pp 1192–1212.
- Schwartz AG, Whitcomb JM, Nyce JW, Lewbart ML, Pashko LL: Dehydroepiandrosterone and structural analogs: A new class of cancer chemopreventive agents. Adv Cancer Res 51:391–424, 1988.
- Orentreich N, Brind JL, Rizer RL: Age changes and sex differences in serum dehydroepiandrosterone sulfate concentrations throughout adulthood. J Clin Endocrinol Metab 59:551–555, 1984.
- Gordon GB, Helzlsouer KJ, Comstock GW: Serum levels of dehydroepiandrosterone and its sulfate and the risk of developing bladder cancer. Cancer Res 51: 1366–1369, 1991.
- Gordon GB, Helzlsouer KJ, Alberg AJ, Comstock GW: Serum levels of dehydroepiandrosterone and dehydroepiandrosterone sulfate and the risk of developing gastric cancer. Cancer Epidemiol Biomarkers Prev 2:33–35, 1993.
- Barrett-Connor E, Khaw K-T, Yen SSC: A prospective study of dehydroepiandrosterone sulfate, mortality, and cardiovascular disease. N Engl J Med 315:1519– 1524, 1986.
- Raineri R, Levy HR: On the specificity of steroid interaction with mammary gland glucose-6-phosphatedehydrogenase. Biochemistry 9:2233–2243, 1970.
- 37. Shantz LM, Talalay P, Gordon GB: Mechanism of in-

hibition of growth of 3T3-L1 fibroblasts and their differentiation to adipocytes by dehydroepiandrosterone and related steroids: Role of glucose-6-phosphate dehydrogenase. Proc Natl Acad Sci USA 86:3852–3856, 1989.

- Pashko LL, Lewbart ML, Schwartz AG: Inhibition of 12-O-tetradecanoylphorbol-13-acetate-promoted skin tumor formation in mice by 16α-fluoro-5-androsten-17-one and its reversal by deoxyribonucleosides. Carcinogenesis 12:2189–2192, 1991.
- Schwartz AG, Perantoni A: Protective effect of dehydroepiandrosterone against aflatoxin B_i-and 7,12-dimethylbenz(*a*)anthracene-induced cytotoxicity and transformation in cultured cells. Cancer Res 35: 2482– 2487, 1975.
- Lee T-C, Lai G-J, Kao S-L, Ho I-C, Wu C-W: Protection of a rat tracheal epithelial cell line from paraquat toxicity by inhibition of glucose-6-phosphate dehydrogenase. Biochem Pharmacol 45:1143–1147, 1993.
- Whitcomb JM, Schwartz AG: Dehydroepiandrosterone and 16α-Br-epiandrosterone inhibit 12-Otetradecanoylphorbol-13-acetate stimulation of superoxide radical production by human polymorphonuclear leukocytes. Carcinogenesis, 6:333-335, 1985.
- 42. Cerutti P: Oxy-radicals and cancer. Lancet 344:862-863, 1994.
- 43. Witzum JL: The oxidation theory of atherosclerosis. Lancet 344:793–795, 1994.
- 44. Jenner P: Oxidative damage in neurodegenerative disease. Lancet 344:796–798, 1994.
- Imlay JA, Linn S: DNA damage and oxygen radical toxicity. Science 240:1302–1309, 1988.
- Stevenson MA, Pollock SS, Coleman CN, Calderwood SK: X-irradiation, phorbol esters, and H₂O₂ stimulate mitogen-activated protein kinase activity in NIH-3T3 cells through the formation of reactive oxygen intermediates. Cancer Res 54:12–15, 1994.
- Kensler TW, Bush DM, Kozumbo WJ: Inhibition of tumor promotion by a biomimetic superoxide dismutase. Science 221:75–77, 1983.
- Babior BM: The enzymatic basis for O₂⁻ production by human neutrophils. Can J Physiol Pharmacol 60:1353–1358, 1982.
- 49. Marletta MA, Yoon PS, Iyengar R, Leaf CD, Wishnok JS: Macrophage oxidation of L-arginine to nitrite and nitrate: Nitric oxide is an intermediate. Biochemistry 27:8706–8711, 1988.
- Sadowski IJ, Wright JA, Israels LG: A permealized cell system for studying regulation of aryl hydrocarbon hydroxylase:NADPH as rate limiting factor in benzo(*a*)pyrene metabolism. Int J Biochem 17:1023– 1025, 1985.
- van Vollenhoven RF, Engelman EG, McGuire JL: An open study of dehydroepiandrosterone in systemic lupus erythematosus. Arthritis Rheum 37:1305–1310, 1994.
- Buffington CK, Pourmotabbed G, Kitabchi AE: Case report: Amelioration of insulin resistance in diabetes with dehydroepiandrosterone. Am J Med Sci 306:320– 324, 1993.

- 53. Levy JC, Burnett M, Manley S, Taylor NF, Burke CW, Turner RC: Dehydroepiandrosterone reduces fasting plasma glucose but not prandial glucose profile or peripheral insulin resistance in type 2 (NIDDM) diabetes. Diabetes 40:345A, 1991 (abstract).
- 54. Schwartz AG, Lewbart ML, Pashko LL: Novel dehydroepiandrosterone analogues with enhanced biological activity and reduced side-effects in mice and rats. Cancer Res 48:4817-4822, 1988.
- 55. Mortola JF, Yen SSC: The effects of oral dehydroepiandrosterone on endocrine-metabolic parameters in postmenopausal women. J Clin Endocrinol Metab 71:696–704, 1990.
- Rao MS, Subbarao V, Yeldani AV, Reddy JK: Hepatocarcinogenicity of dehydroepiandrosterone in the rat. Cancer Res 52:2977–2979, 1992.
- Ratko TA, Detrisac CJ, Mehta RG, Kelloff GJ, Moon RC: Inhibition of rat mammary gland chemical carcinogenesis by dietary dehydroepiandrosterone or a fluorinated analogue of dehydroepiandrosterone. Cancer Res 51:481–486, 1991.
- Pashko LL, Schwartz AG: Anti-hyperglycemic effect of the dehydroepiandrosterone analogue 16α-fluoro-5-androsten-17-one in diabetic mice. Diabetes 42: 1105–1108, 1993.